BMB471: ADVANCED BIOCHEMISTRY LABORATORY SYLLABUS SS23 Edition

Table of Contents

Instructors	2
Books Used in Biochemistry and Molecular Biology 471	2
Texts	
Handbooks	2
Objectives	3
Organization of Course	3
Computer Resources	3
Course Schedule & Due Dates	
Pre-lab problem sets	
Course reading assignments for Monday lectures	5
Bi-weekly quizzes	5
Grading	5
Challenging Grades	
COVID-19 Safety precautions	6
Laboratory Safety	
General Laboratory Rules	
In Case of an Accident	
Academic Integrity	9
Student Rights and Responsibilities	10
Teaching Labs Floor Plan	10
Emergency Evacuation Instructions	11

Instructors

Faculty & Staff	Email	Office	Phone	Office Hours**
R. Michael Garavito*	garavito@msu.edu	513 BCH	(517) 355-9724	Thursday 4-5 pm
Björn Hamberger	hamberge@msu.edu	3230 MPS	(517) 884-6964	Thursday 4-5 pm
Allan TerBush	terbusha@msu.edu	113 BCH	(517) 355-3971	Monday 10 am -12 pm

^{*}Course coordinator

^{**}We are certainly open to setting individual appointments upon request (in person or via Zoom)

Teaching Assistants	Email
Ashley Caughell	caughel1@msu.edu
Larissa Ford	fordlari@msu.edu
Priyam Patel	patelp60@msu.edu
Winston Diep	diepwins@msu.edu

Books Used in Biochemistry and Molecular Biology 471

<u>Texts</u>

Required texts:

- Advanced Biochemistry Laboratory, 2023 ed.
- Advanced Biochemistry Laboratory Supplement, 2023 ed.

Recommended texts:

- Segel, I.H., Biochemical Calculations, 2nd ed., J. Wiley & Sons, Inc., 1976.
- Day, R.A., *How to Write and Publish a Scientific Paper*, 6th or 7th ed. Oryx Press, 2006.
- Boyer, R.F., *Modern Experimental Biochemistry*, 3rd ed., Addison Wesley Pub. Co., 2000.
- Voet, D. and Voet, J.G., Biochemistry, 2nd edition, J. Wiley & Sons, N.Y., N.Y., 1995.
 Available in the Teaching Labs and Room 105 Biochemistry:

Handbooks

- Handbook of Chemistry & Physics, CRC Press (various editions, a new edition is published each year)
- The Merck Index, Merck & Co. (various editions)
- Dawson, R.M., Elliott, D.C., Elliott, W.H., and Jones, K.M., *Data for Biochemical Research*, 3rd ed., Clarendon Press, Oxford, 1986.
- Sax, N.I., *Dangerous Properties of Industrial Materials*, 4th or 5th ed., Van Nostrand Reinhold Co., 1975 (4th), 1979 (5th)
- Rayburn, S.R., The Foundations of Laboratory Safety, Springer-Verlag, 1990.

Objectives

The subject areas for the course are weak acids and bases, spectrophotometry, protein purification, and enzyme activity assays. The objectives for the course are for students to:

- Develop the ability to design logical experiments given specific experimental objectives but only general procedures, perform these experiments successfully and independently, properly interpret the data, and clearly present the data in writing.
- Learn foundational biochemical laboratory methodology and techniques.
- Further develop:
 - o quantitative laboratory skills.
 - the ability to keep a clear and complete lab notebook.
 - the ability to analyze and interpret laboratory data.
 - o skill in writing scientific laboratory reports.
- Understand the concepts on which the laboratory experiments are based and related concepts (largely covered in lecture and homework problems).

Organization of Course

The course consists of one 50-minute lecture, and one 5-hour laboratory period per week. The lectures will be in-person in room 101 of the Biochemistry at 12:40 pm EST and are **mandatory**. Biweekly quizzes will be given in D2L to test your understanding of the course material. You will have 20 minutes to complete each quiz.

The course will have a semi-open lab format; students may leave and return to the lab rooms at any time. The labs (113 & 117 Biochemistry) will be open Tuesday through Friday from 12:30 to 5:30 p.m., and students may work only on their scheduled lab day.

Each student will work individually in Units A, B, C, and E. For Unit D, students will work in groups of two. When working in pairs, partners only collaborate on the experimental (data acquisition) portions of the labs; all notebook preparation, data analysis/post-lab notebook entries, and lab reports are to be done individually. Unit E is considered a practical examination for the course. Students are expected to perform experimental work each lab period. Except in cases of excused absence, students who do not participate in data collection during the lab period are not entitled to the data collected by others.

Computer Resources

The Teaching Lab has computers with internet access that may be used during scheduled laboratory hours. We will be using the Desire2Learn (D2L) system for posting announcements and course materials as well as LON-CAPA for pre-lab problem sets (according to the schedule posted on Page 5), and submission of your laboratory reports. It is recommended that students have access to 25 Mbps download speed for watching video content from D2L.

Course Schedule & Due Dates

Lab Period	Date	Experimental Work	Due Dates	LON-CAPA ¹	Bi-weekly Quiz ²
1	Jan. 9-13	Lab introduction; check-in; safety tour; student practice with spectrophotometers and chart recorders			
2	Jan. 16-20	Unit A - purity of light & wavelength accuracy		PS#1 Unit A (day 1)	
3	Jan. 23-27	Unit A - absorbance accuracy & linearity, recorder use		PS#2 Unit A (day 2)	
4	Jan. 30 - Feb. 3	Unit B - buffer preparation (titration curve); pH measurements	Jan. 31 - Feb 3: Notebooks ³ Unit A	PS#3 Unit B (day 1)	Quiz 1
5	Feb. 6-10	Unit B - effects of salt & dilution on pH of a buffer		PS#4 Unit B (day 2)	
6	Feb. 13-17	Unit C – AAT enzyme assays and Lowry protein assay	Feb. 14-17: Notebooks Unit B	PS#5 Unit C	Quiz 2
7	Feb. 20-14	Unit D - AAT heat denaturation and AAT assays	Feb. 21-24: Notebooks Unit C	PS#6 Unit D (day 1)	
8	Feb. 27 - Mar. 3	Unit D – large-scale (NH ₄) ₂ SO ₄ precipitation, and AAT assays	Mar. 3: Report I ⁴ Units A-C	PS#7 Unit D (day 2)	Quiz 3
	Mar. 6-10	Labs Canceled - Spring Break			
9	Mar. 13-17	Unit D - CM-sephadex column chromatography - AAT and Lowry protein assay of AAT fractions	Mar. 14-17: Notebooks D1-D2	PS#8 Unit D (day 3)	
10	Mar. 20-24	Unit D - SDS-PAGE of AAT fractions Unit E - buffer/ethanol preparation, yADH assay testing		PS#9 Unit D (day 4) Unit E (day 0)	Quiz 4
11	Mar. 27-31	Unit E - [yADH], [NAD ⁺], yADH stability	Mar. 28-31: Notebooks D3-D5		
12	Apr. 3-7	Unit E - approximate $K_m \& V_{max}$, stability	Apr. 7: Report II Unit D		Quiz 5
13	Apr. 10-14	Unit E - precise valid range, stability	Apr. 11-14: Notebooks E0-E3	PS#10 Unit E (day 3)	
14	Apr. 17-21	Unit E - precise $K_m \& V_{max}$ and inhibition, stability		PS#11 Unit E (day 4)	
15	Apr. 24-28	Lab check-out	Apr. 24: Notebooks ⁵ E3-E6 Apr. 28: Report III Unit E		Quiz 6

- 1. LON-CAPA Problem Sets (PS) are due **Mondays at 11:30 pm EST**. Each problem set opens 1 week before the due date.
- 2. Quizzes will be given through the D2L course page and open at 1:30 pm the Monday of the week listed on the course schedule and remain available through 9 pm.
- 3. Lab notebooks are due and submitted to their corresponding D2L assignment dropbox by **12:30 pm EST** on the day of your lab section in the weeks noted above.
- 4. Reports must be submitted to their corresponding D2L assignment dropbox

REPORTS ARE DUE BY 12:30 pm EST. - <u>LATE REPORTS RECEIVE NO CREDIT</u>

5. Final lab notebooks (E3-E6) are due Monday, Apr. 24 at 12:30 pm EST.

Pre-lab problem sets

Short problem sets will be available through LON-CAPA. The problems will test your understanding of the experiments and prepare you for pre-lab and post-lab calculations/data analysis for experiments performed the following week. The assignments are due 11:30 pm EST on Mondays. The schedule for distribution and due dates of the problems is as follows:

Course reading assignments for Monday lectures.

Date	Subject	Reading
Jan. 9, 16, 23	Spectrophotometry, including Beer's and Lambert's laws	 Experimental Biochemistry Supplement, Chapter 1 & 3. Segel, I.H. <i>Biochemical Calculations</i>, 2nd ed., pp. 324 - 333
Jan. 30 Feb. 6	Buffers, activity, and related subjects	 Experimental Biochemistry Supplement, Chapter 2. Segel, I.H., Biochemical Calculations, 2nd ed. pp. 1-83.
Feb. 13	Assay of enzymes & protein	1. Experimental Biochemistry Supplement, Chapter 4. 2. Segel, I.H., <i>Bioch. Calcs.</i> , 2 nd ed. pp. 208-214, 291-92, 333-46.
Feb. 20	Protein purification	 Experimental Biochemistry Supplement, Chapter 7. Segel, I.H., Bioch. Calcs., 2nd ed. pp. 287-290
Feb. 27 Mar. 13 Mar. 20	Protein purification Chromatography SDS-PAGE	 Experimental Biochemistry Supplement, Chapter 5. Experimental Biochemistry Supplement, Chapter 6.
Mar. 27 - Apr. 24	Chemical and Enzyme Kinetics	 Experimental Biochemistry Supplement, Chapter 8. Segel, I.H., Biochemical Calculations, 2nd ed., pp. 208-212 (top), 214-219, 221 (bottom)-236 plus problem 4.9 on page 237, 246-250, 252.

Bi-weekly quizzes

Some of the main course goals are for you to be able to analyze and interpret laboratory data for meaningful biochemical conclusions. Therefore, the quizzes will focus on your understanding of the quantitative and conceptual aspects of the experimental work, the material covered in lectures, the assigned readings, and pre-lab problem sets. Problems of the type encountered in the experimental work and in the pre-lab problems may be on the quizzes. You should also expect some short-answer questions focusing on the theory of your experiments and the molecular interactions involved in them.

Have a calculator and transparent ruler available for when you take the bi-weekly quizzes. Once a quiz is begun, you will have 20 minutes to complete it. After the 20 minute time limit (unless otherwise noted by the instructors), no further answers will be allowed.

Grading

Final grades will be based on a weighted grading scale as described below:

Laboratory notebooks	40%
Laboratory reports	30%
Quizzes	20%
Prelab problem sets	10%

Students earning 90% or more of the total points will receive a grade of 4.0. The dividing line between 2.5 and 2.0 grades will be approximately 70% of the total points. The range between 90% and 70% will be divided into approximately equal parts for assigning grades of 3.5, 3.0, and 2.5. The dividing line between 1.0 and 0.0 grades will be about 50%. A passing grade will be awarded to any student who completes and submits all Problem Sets, Lab Reports and Notebooks on time, participates in all laboratory sessions (except for excused absences), and earns at least 50% of the points.

Note: Reports are <u>due by 12:30 pm EST.</u> on the day listed in the schedule. <u>Reports submitted after the due time/date will not receive any credit</u>. Electronic templates will be provided for you to make your reports from. Submit your electronic reports to their corresponding D2L assignment dropbox.

Note: Notebooks are <u>due at 12:30 pm EST on the day of your lab section</u> (except the last grading; <u>see schedule</u>) the week listed in the schedule. Late notebooks will incur a <u>25% per 24-hour period penalty</u> (including weekends) and may have a delay in when feedback and grades are published. Notebooks submitted more than 3 days late will receive no credit. They will be corrected with feedback provided. Additionally, objects (data tables, graphs, chart tracings, etc) placed into incorrect sections of the notebooks will result in a 50% point deduction for the affected objects. There is a very thorough description of notebook organization in the Notebook section of the course pack (pages 3-5).

Challenging Grades

If you believe an assignment was graded incorrectly, you have 2 weeks from the time grades and feedback are published to email Dr. TerBush to challenge the grade. You must include a listing of the criteria that you believe are graded incorrectly and a description of where in the assignment you earned those points. Grade challenges after 2 weeks will not be considered.

COVID-19 Safety precautions

The COVID-19 virus is extremely contagious and is believed to spread mainly from person-to-person contact. Due to the continued presence of the novel coronavirus, various policies have been implemented for in-person learning. This section of the syllabus discusses the policies and procedures we will use in class.

This class abides by all principles, guidelines, and requirements detailed in the MSU Community Compact. Specifically, all participants in the class will adhere to the following:

Personal Hygiene. All students must maintain proper hygiene and health practices, including:

- Washing hands frequently with soap and water or, if soap is unavailable, using hand sanitizer with at least 60% alcohol
- Routinely cleaning and sanitizing living spaces and/or workspace
- Using the bend of the elbow or shoulder to shield a cough or sneeze

Adherence to Signage and Instructions. Students will (a) look for instructional signs posted by MSU or public health authorities, (b) observe instructions from MSU or public health authorities that are emailed to my "msu.edu" account, and (c) follow those instructions.

Self-Monitoring. Students will self-monitor for flu-like symptoms (for example, cough, shortness of breath, difficulty breathing, fever, sore throat or loss of taste or smell). If a student experiences any

flu-like symptoms, they will stay home and contact a health care provider to determine what steps should be taken.

Exposure to COVID-19. If a student is exposed to someone who is ill or has tested positive for the COVID-19 virus, they will contact the MSU COVID-19 Triage line (1-888-958-2678) and follow the instructions for how to proceed.

Compliance and reporting. Those who come to MSU facilities must commit to the personal responsibility necessary for us to remain as safe as possible, including following the specific guidelines outlined in this syllabus and provided by MSU more broadly (see below). There may be times when action will be necessary to reinforce expectations. If you do not wear appropriate face coverings (see MSU's guidelines) or do not wear your face covering appropriately (i.e., over your mouth and nose) you will be asked to correct the situation or leave the facility. In addition, MSU will utilize the processes already in place to respond to any issues of noncompliance with standards established for the health and safety of our community. For classroom disruptions or issues, the responses and processes that have been used previously remain the first line of action. If necessary, the student conduct system will be the avenue used to adjudicate student disciplinary situations.

Laboratory Safety

For the welfare of fellow students, and for your own well-being, each student is expected to follow a set of accepted laboratory precautions. To ensure that each student understands these guidelines, students are expected to <u>sign a consent form</u>. The form will be provided on the first day of class. In the lab:

- Protective eyewear must be worn at all times (safety glasses or face shields). No exceptions to this policy will be made.
 - A few times throughout the semester, you will need chemical splash goggles when working with a strong acid or base. You will be informed of these lab periods ahead of time.
- Proper clothing (full-length pants, close-toed shoes, full-length shirts (no bare mid-drifts)) must be worn to lab. Put simply, no skin may be exposed beneath the waist. Students with improper attire will be asked to leave and come back with proper clothing.
- Gloves must be worn when working with hazardous chemicals (Units B and D). We will supply suitable gloves for this purpose.
- Do not eat, drink, or apply cosmetics at any time while in the laboratory. **Do not place food** wrappers, beverage cups, or other food-related waste in the trash cans in the lab.
- Mouth pipetting is not permitted.
- Do not sit on the lab benches.
- Use chemicals with high vapor pressure only in the hood.
- Handle and dispose of hazardous chemicals properly. Disposal containers are provided.
- When chemicals are spilled, they should be wiped or swept up or both as soon as possible. If the spillage is large, immediately notify an instructor. Sponges and brushes are provided and should be used to keep your work area clean.
- Be aware of objects that can burn or give electrical shocks.
- Do not use open flames near flammable chemicals.
- In general, be alert to possible hazards from chemicals, glassware, electrical connections, and flammable solvents. Read labels and observe suggested precautions.
- Sweep up broken glassware as soon as possible and deposit it in the labeled container set aside for this purpose.
- Never work alone.
- All solutions that you prepare must be labeled with date, your last name, and the type and concentration of the reagent. Unmarked solutions will be disposed.
- Wipe your lab bench with a damp sponge at the end of each lab period.
- Be familiar with location and use of safety items location of safety showers, eyewash stations, fire blankets, fire extinguishers, medical kit. Review the lab floor plan and evacuation routes on page 11 before coming to the first lab period.

General Laboratory Rules

 Used glassware from the glassware cabinets must be rinsed with tap water and placed on the designated cart after all tape has been removed. Ink writing on this glassware need not

- be removed. Glassware from the shared bench drawers must be rinsed with tap water, then rinsed several times with de-ionized water and returned to the shared drawer.
- Items stored in the cold room or freezer boxes should be removed and disposed when they are no longer needed.
- The instructors will make every effort to keep equipment in good working order. It is your responsibility to read and understand instructions for the use of equipment. Do not turn on an instrument until you have done this. Do not hesitate to ask questions of your instructor after you have read the instructions. If any equipment malfunction is noted, report this immediately to an instructor.
- Be considerate of others. Clean up after yourself after using any piece of equipment. All equipment, especially balances, should be left ready for immediate use by another student.

In Case of an Accident

- Report all injuries, even minor ones, to an instructor immediately.
- In case of even minor laboratory accidents, you should go to Olin Health Center for treatment. Your instructor will provide an accident report form; a copy of this form must be taken to the Center.
- For any chemicals splashed in the eye, hold the eye open and flush immediately with cold water by using the eye wash. Flush for at least 5 minutes and call for help from an instructor.
- For chemicals spilled on the skin or splashed into the mouth, again, flush with large amounts of cold water for 5 or more minutes. Call for an instructor.
- For severe bleeding, apply pressure and call for help from an instructor.
- For burns, flush with cold water and contact an instructor.
- Note: Injury to students in the laboratory is not covered by University insurance. Students must provide their own insurance coverage.

Academic Integrity

Academic integrity is central to the educational process and is expected throughout this course. We will teach you accepted standards of intellectual honesty with respect to the performance of experiments, the processing of data, and the reporting of results. We expect you to follow these standards. Particularly be aware of when you must work independently. Credit will not be given to work obtained through collaboration when independent effort is required. Students should be familiar with the student handbook, which governs all student behavior.

Please see the following additional expectations below:

Each student is to work completely independently in the processing of their data, as well as in the writing of the reports. Students are welcome to discuss their data with each other, with instructors, or with others to achieve a full understanding of them before processing them. The exception to open discussion: In the units where a coworker collects some of your data, you may not discuss the processing of the data or the preparation of the report with your coworker in any way. Discussions about the data and experiment with anyone else are still encouraged. Plagiarism of any kind will not be tolerated. Penalties for plagiarism will range from a severe point deduction to a score of zero for the assignment, and as such will be regarded as a "penalty grade". In accordance with university policy, a report will be forwarded to the academic dean, which will become a part of the student's academic record. In addition, falsification or fabrication of any

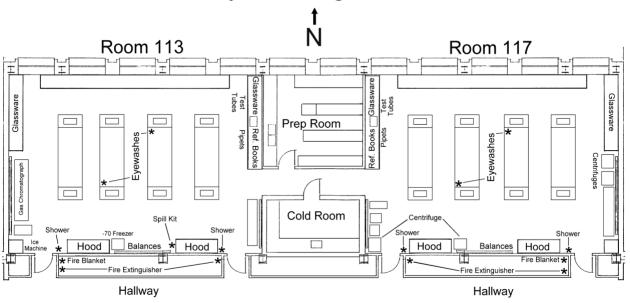
laboratory data will result in a failing grade in the class and a report will be forwarded to the academic dean.

Student Rights and Responsibilities

As students, you have a range of support and information options available to discuss actions or activities related to their academic, personal or professional lives at MSU. These rights and resources are detailed on the ombudsperson's website.

Teaching Labs Floor Plan

Biochemistry Teaching Labs - Floor Plan



Emergency Evacuation Instructions

In Case of an Emergency

Fire: Exit room and go left (East) to EXIT and move away from building Tornado: Exit room and go left (East) and downstairs to middle of building

